



Hematopoietic Stem Cell Transplantation Successfully Treats *CD40LG* Duplication

Di Sun¹ · Carole Le Coz¹ · Nancy Bunin² · Neil Romberg^{1,3,4}

Received: 29 April 2021 / Accepted: 8 June 2021 / Published online: 22 June 2021
© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2021

To the editor:

CD40LG encodes CD40L, a tightly regulated T-cell coactivation receptor, that binds CD40 on antigen presenting cells such as dendritic cells, macrophages, and B cells. Physiologic CD40L induction promotes T-cell apoptosis and B-cell class switching [1–3]. Supraphysiologic CD40L expression promotes autoantibody production and portends poor prognoses in patients with lymphopenia-associated autoimmune diseases [3].

Recently, we described a boy with infantile-onset Evans syndrome, massive splenomegaly, and IgM deficiency who maternally inherited a 240 kb chromosome X microduplication (ChrX:135,539,461–135,780,648) encompassing *CD40LG* and its regulatory elements [4]. His T lymphocytes, which overexpressed CD40L upon induction, demonstrated accelerated cell death and exuberantly promoted B-cell immunoglobulin class-switching in vivo and in vitro [4]. Although his autoimmune manifestations were successfully managed with cyclosporine, which normalized CD40L expression by inhibiting nuclear factor of activated T cells (NFAT), his T-cell lymphopenia was insidiously progressive. At eight years old, his peripheral blood CD3⁺ T-cell concentration fell to 334 cells/ μ L (134 CD4⁺ cells/ μ L and 154 CD8⁺ cells/ μ L; Supplemental Fig. 1) and he was referred for hematopoietic stem cell transplantation (HSCT).

After myeloablative conditioning with 3.2 mg/kg/dose busulfan (day –9 to –4), 30 mg/m²/dose fludarabine (day –6

to –3, –2), 3 mg/kg/dose anti-thymocyte globulin (day –5 to –3), and 30 mg/m²/dose cyclophosphamide (day –3 to –2), the patient received a 9/10 matched unrelated donor T-cell receptor α/β +/CD19⁺ depleted peripheral stem cell transplant. His only transplant-related adverse event was BK virus cystitis which resolved with aggressive hydration. During conditioning, the patient experienced total and durable resolution of all autoimmune manifestations. Other than standard graft-versus-host disease prophylaxis with mycophenolate mofetil for 45 days post-transplant, he required no further immunosuppressant therapies. One month after HSCT, 99% of the patient's myeloid, B-cell, and NK lineage cells were donor-derived (Fig. 1A). In contrast, T-cell-mixed chimerism was significant. A total of ~40% of T cells were of donor origin a month after transplantation, but the donor fraction slowly increased to 93% by 18 months (Fig. 1A). Despite rapid numerical naive B-cell reconstitution (Supplemental Fig. 1), the patient remained immunoglobulin replacement therapy-dependent after transplantation. In contrast to the excessive in vivo class-switching observed pre-HSCT, circulating class-switched memory B cells (0–1 CD19⁺CD27⁺IgM⁻ cells/ μ L) and all immunoglobulin isotypes were scarce during the post-HSCT period.

To functionally assess correction of the patient's immune defect by HSCT, we subjected patient CD4⁺ T cells cryopreserved before and 18 months after transplant to a series of functional in vitro studies. Compared to pre-transplant patient CD4⁺ T cells, which overexpressed CD40L upon activation, CD40L induction by post-transplant patient cells was similar to healthy donor (HD) counterparts (Fig. 1B). Similarly, while 16.7% of naïve HD B cells co-cultured with pre-HSCT patient CD4⁺ T cells class-switched to IgG, only 10.8% and 7.8–11.8% of naïve HD B cells IgG class-switched when co-cultured with post-transplant patient CD4⁺ T cells or HD CD4⁺ T cells, respectively (Fig. 1C). Finally, cell death, measured with annexin V and 7-AAD co-staining after culturing live-sorted CD4⁺ cells for 48 h in culture, was similar

✉ Neil Romberg
rombergn@chop.edu

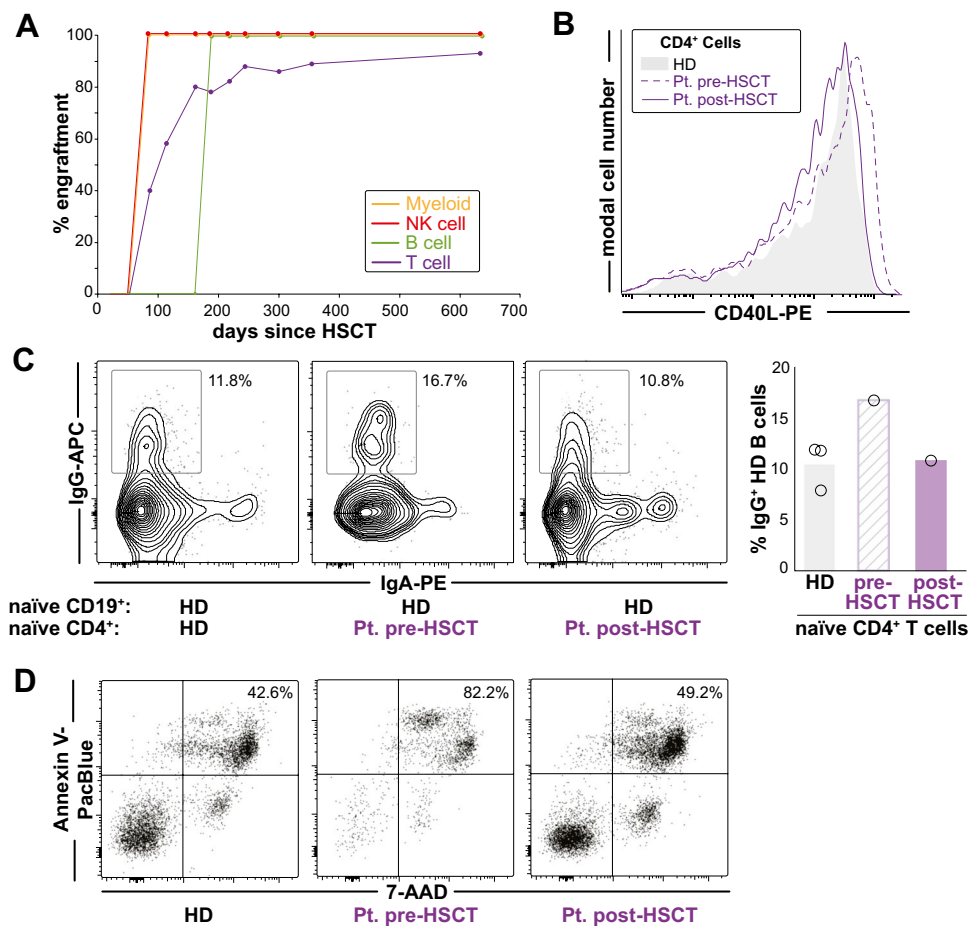
¹ Division of Immunology and Allergy, Children's Hospital of Philadelphia, Philadelphia, PA 19104, USA

² Division of Oncology, Children's Hospital of Philadelphia, Philadelphia, PA 19104, USA

³ Department of Pediatrics, Perelman School of Medicine, Philadelphia, PA 19104, USA

⁴ Abramson Research Center, Children's Hospital of Philadelphia, Room 1216C, Philadelphia, PA 19104, USA

Fig. 1 Lineage engraftment and in vitro functional studies after hematopoietic stem cell transplantation (HSCT). **A** Lineage-specific engraftment following HSCT. **B** CD40L expression on healthy donor (HD) CD4⁺ T cells (mean fluorescence intensity, MFI 4529) and patient (Pt.) CD4⁺ T cells either pre- or post-HSCT (MFIs 5850 and 4502, respectively). Assessments were made 8 h after phorbol 12-myristate-13-acetate (PMA) induction. **C** Representative flow cytometry plots of IgG expression on HD naïve B cells following a 7-day co-culture with anti-CD3/CD28-activated CD4⁺CD45RO⁻ T cells from either a different HD, the patient pre-HSCT, or the patient post-HSCT. Bar graph depicts IgG⁺ HD B-cell frequencies after co-culture with CD4⁺ T cells from three HD controls (2 male, 1 female) and the patient pre- and post-HSCT. **D** Annexin V and 7-AAD staining of CD4⁺ T cells after 48 h in anti-CD3/CD28 antibody activating conditions. At time zero, all cells were verified viable through sorting



in HD and post-HSCT patient cells (42.6% and 49.2%, respectively) (Fig. 1D). In contrast, nearly all (82.2%) pre-HSCT patient cells died in culture.

The clinical phenotype associated with CD40L over-expression—autoantibody-mediated autoimmune disease and lymphopenia—illustrates the two seemingly disparate consequences of CD40/CD40L engagement: B-cell activation or T-cell death. Although our *CD40LG*-duplicated patient's autoimmune disease was responsive to CD40L-modulating cyclosporine, his progressive lymphopenia required more definitive cellular therapy. HSCT works especially well when donor cells enjoy a competitive survival advantage over host cells. Although *CD40LG*-duplicated T cells demonstrate accelerated cell death, our patient's donor T-lineage engraftment was delayed. As thymic epithelial architecture is disrupted in CD40L-over-expressing transgenic mice, it may be that our patient's thymus required a normalization period before it could fully accommodate donor thymocyte development [5]. Regardless, the patient's clinical course and normalization of T-cell function post-HSCT reaffirm CD40L over-expression's role in disease pathogenesis and highlights that *CD40LG* duplication is a transplantable condition.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10875-021-01085-5>.

Author Contribution C.L.C. and N.R. planned the experiments. D.S. and C.L.C. performed the experiments and completed data analysis. D.S. and N.R. wrote the manuscript, and C.L.C. and N.B. edited the manuscript. N.B. and N.R. provided patient clinical data. All authors approve of the manuscript.

Funding N.R. receives salary support from the Jeffrey Modell Foundation. D.S. is funded by NICHD-T32HD043021.

Data Availability Not applicable.

Code Availability Not applicable.

Declarations

Consent for Publication All authors have reviewed the manuscript and consented to publication.

Conflict of Interest The authors declare no competing interests.

References

1. Blair PJ, Riley JL, Harlan DM, Abe R, Tadaki DK, Hoffmann SC, et al. Cd40 ligand (Cd154) triggers a short-term Cd4+ T cell activation response that results in secretion of immunomodulatory cytokines and apoptosis. *J Exp Med.* 2000;191:651–60.
2. Higuchi T, Aiba Y, Nomura T, Matsuda J, Mochida K, Suzuki M, et al. Cutting edge: ectopic expression of CD40 ligand on B cells induces lupus-like autoimmune disease. *J Immunol.* 2002;168:9–12.
3. Karnell JL, Rieder SA, Ettinger R, Kolbeck R. Targeting the CD40-CD40L pathway in autoimmune diseases: humoral immunity and beyond. *Adv Drug Deliv Rev.* 2019;141:92–103.
4. Le Coz C, Trofa M, Syrett CM, Martin A, Jyonouchi H, Jyonouchi S, et al. CD40LG duplication-associated autoimmune disease is silenced by nonrandom X-chromosome inactivation. *J Allergy Clin Immunol.* 2018;141:2308–2311.e7.
5. Dunn RJ, Luedecker CJ, Haugen HS, Clegg CH, Farr AG. Thymic overexpression of CD40 ligand disrupts normal thymic epithelial organization. *J Histochem Cytochem.* 1997;45:129–41.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.